

The Roles of Phasic and Tonic Dopamine in Tic Learning and Expression

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ABSTRACT

Tourette syndrome (TS) prominently involves dopaminergic disturbances, but the precise nature of those disturbances has remained elusive. A substantial body of empirical work and recent computational models have characterized the specific roles of phasic and tonic dopamine (DA) in action learning and selection, respectively. Using insights from this work and models, we suggest that TS involves increases in both phasic and tonic DA, which produce increased propensities for tic learning and expression, respectively. We review the evidence from reinforcement-learning and habit-learning studies in TS, which supports the idea that TS involves increased phasic DA responses; we also review the evidence that tics engage the habit-learning circuitry. On the basis of these findings, we suggest that tics are exaggerated, maladaptive, and persistent motor habits reinforced by aberrant, increased phasic DA responses. Increased tonic DA amplifies the tendency to execute learned tics and also provides a fertile ground of motor hyperactivity for tic learning. We review evidence suggesting that antipsychotics may counter both the increased propensity for tic expression, by increasing excitability in the indirect pathway, and the increased propensity for tic learning, by shifting plasticity in the indirect pathway toward long-term potentiation (and possibly also through more complex mechanisms). Finally, we review evidence suggesting that low doses of DA agonists that effectively treat TS decrease both phasic and tonic DA, thereby also reducing the propensity for both tic learning and tic expression, respectively.

Keywords: Basal ganglia, Computational psychiatry, Phasic dopamine, Tics, Tonic dopamine, Tourette syndrome

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Tourette syndrome (TS) has long been thought to involve dopaminergic disturbances (1,2). Despite substantial work investigating the dopaminergic system in TS (3,4), however, the nature of these disturbances has remained controversial. As we review in a recently submitted article (T.V. Maia, Ph.D., and V.A. Conceição, M.S., unpublished review, February 2017), existing hypotheses explain only restricted subsets of findings and, in some cases, are contradicted by substantial evidence. In that article, we show that the hypothesis that TS involves striatal dopaminergic hyperinnervation provides a parsimonious and integrated explanation for the various findings from studies of the dopaminergic system in TS. This dopaminergic hyperinnervation creates a hyperdopaminergic state, which explains why all medications that are effective for TS reduce dopamine (DA) neurotransmission (Maia and Conceição, unpublished review, February 2017). This hyperinnervation likely causes both phasic and tonic DA to be increased. Building on computational ideas about the roles of phasic and tonic DA in action learning and selection, respectively (5–7), in the present article we show computationally that increased phasic and tonic DA cause increased propensities to learn and express tics, respectively.

Phasic DA represents reinforcement prediction errors (PEs) (8,9), which support model-free, habit learning (10). We suggest that aberrant, exaggerated phasic DA signals abnormally reinforce motor behaviors, thereby causing tic learning.

In other words, we suggest that tics are exaggerated, maladaptive, and persistent motor habits learned through aberrant phasic DA signals.

Tonic DA invigorates action (11–13) and increases the expression of previously learned behaviors (5). We suggest that increased tonic DA promotes tic expression by amplifying the tendency to execute learned tics; in addition, it increases the overall tendency for action, thereby creating a fertile ground for tic development.

COMPUTATIONAL ROLES OF PHASIC AND TONIC DA

Phasic DA bursts represent positive PEs (8,9) that drive action learning in the basal ganglia, strengthening corticostriatal synapses onto direct-pathway (Go) medium spiny neurons (MSNs), which represent the positive or Go value (G) of actions, and weakening corticostriatal synapses onto indirect-pathway (NoGo) MSNs, which represent the negative or NoGo value (N) of actions (5–7,14–16) (see Box 1). Exaggerated, and possibly abnormally timed, phasic DA signals therefore may cause exaggerated, aberrant learning of motor behaviors, thereby producing tics.

Whereas phasic DA bursts elicit transient DA concentrations ranging from 10 nM to more than 1 μ M (17), tonic DA concentrations are in the low nanomolar range (18). In the striatum, most D_1 and D_2 receptors are in low- and high-affinity states, respectively (19), which are sensitive to micromolar and

Box 1. Computational Roles of Dopamine

Phasic Dopamine and Action Learning

Dopamine (DA) signals reinforcement prediction errors (PEs), which are the difference between the obtained and expected reinforcement (8,9). The reinforcement that a person expects to get in a given situation, s , is usually called the value of that situation and is represented by $V_t(s)$ (where the time subscript, t , indicates that this value can change over time). The PE, δ_t , is then given by

$$\delta_t = r_t - V_t(s_t),$$

where r_t and s_t are the reinforcement received and the state at time t , respectively.

PEs are useful because they can be used both to improve the estimate of the values of situations and to learn which actions to take in each situation. In the actor-critic model (143), these two processes are implemented in the critic and the actor, respectively, which map onto distinct neural substrates (8,144). The critic updates the values of the situation, s , using the following equation:

$$V_{t+1}(s_t) = V_t(s_t) + \alpha_C \delta_t,$$

where α_C is a parameter between 0 and 1 that represents the critic's learning rate. This equation ensures that the estimate of $V(s_t)$ is moved somewhat in the direction of r_t , making $V(s_t)$ an exponential recency-weighted average of past reinforcements (8). The actor similarly uses the PE to update its preference, $\rho(s_t, a_t)$, for taking the action that was executed, a_t , in situation s_t :

$$\rho_{t+1}(s_t, a_t) = \rho_t(s_t, a_t) + \alpha_A \delta_t,$$

where α_A is a parameter between 0 and 1 that represents the actor's learning rate. This equation ensures that actions that result in positive and negative PEs are strengthened and weakened, respectively.

The basal ganglia contain partly opposing direct (Go) and indirect (NoGo) pathways (6,145) that can be conceptualized as implementing two opponent actors: one, the direct (Go) pathway, that stores the positive values of actions, and another, the indirect (NoGo) pathway, that stores the negative values of actions (5,7). DA affects plasticity in corticostriatal projections to D_1 and D_2 medium spiny neurons (MSNs) of the direct and indirect pathways, respectively, in opposite ways: broadly speaking, DA stimulation of D_1 and D_2 receptors promotes long-term potentiation (LTP) and long-term depression (LTD) in D_1 and D_2 MSNs, respectively, and lack of DA stimulation promotes LTD and LTP in D_1 and D_2 MSNs, respectively (14,15). Therefore, PEs affect learning in Go and NoGo pathways in opposite ways:

$$G_{t+1}(s_t, a_t) = \begin{cases} G_t(s_t, a_t) + \alpha_{G,LTP} \delta_t, & \text{if } \delta_t \geq 0 \\ G_t(s_t, a_t) + \alpha_{G,LTD} \delta_t, & \text{if } \delta_t < 0 \end{cases}$$

and

$$N_{t+1}(s_t, a_t) = \begin{cases} N_t(s_t, a_t) - \alpha_{N,LTD} \delta_t, & \text{if } \delta_t \geq 0 \\ N_t(s_t, a_t) - \alpha_{N,LTP} \delta_t, & \text{if } \delta_t < 0 \end{cases}$$

where $G(s_t, a_t)$ and $N(s_t, a_t)$ represent the striatal Go and NoGo values, respectively, of the association between state s_t and action a_t (which are stored in corticostriatal synapses onto D_1 and D_2 MSNs, respectively) and are constrained to be greater than or equal to 0. The parameters $\alpha_{G,LTP}$, $\alpha_{G,LTD}$, $\alpha_{N,LTP}$, and $\alpha_{N,LTD}$ are learning rates between 0 and 1, with the subscripts G and N indexing plasticity in Go and NoGo MSNs, respectively, and the subscripts LTP and LTD representing LTP and LTD plasticity, respectively.

Tonic Dopamine and Action Selection

Actions are selected on the basis of the difference between their Go (G) and NoGo (N) values (5,6). A common function for such selection is the softmax (146), which in this case relates the difference between G and N for each action to the probability, $P_t(a|s_t)$, of selecting that action in current situation s_t :

$$P_t(a|s_t) = \frac{e^{\beta [G_t(s_t, a) - N_t(s_t, a)]}}{\sum_{a_i} e^{\beta [G_t(s_t, a_i) - N_t(s_t, a_i)]}}$$

where β is a gain parameter that controls the degree of randomness in action selection, and a_i ranges over all available actions in situation s_t .

DA at the time of action selection modulates the gain (i.e., the excitability) of Go and NoGo MSNs in opposite directions: broadly speaking, DA increases the gain of Go MSNs (i.e., it makes Go MSNs more excitable) through its stimulatory action on D_1 receptors, and it decreases the gain of NoGo MSNs (i.e., it makes NoGo MSNs less excitable) through its inhibitory action on D_2 receptors (5,147). Thus, instead of a single gain, β , that applies to both pathways, there are two gains, β_G and β_N , that apply to the Go and NoGo pathways, respectively:

$$P_t(a|s_t) = \frac{e^{\beta_G G_t(s_t, a) - \beta_N N_t(s_t, a)}}{\sum_{a_i} e^{\beta_G G_t(s_t, a_i) - \beta_N N_t(s_t, a_i)}}$$

These two gains are modulated by DA in opposite directions (5) and depend on the total amount of DA, ρ , present during action selection, which consists of the sum of tonic DA, γ , and phasic DA signals due to any PEs, δ , that occur shortly before action selection (7):

$$\rho = \gamma + \delta.$$

A convenient mathematical formulation to capture the opposite modulation of β_G and β_N by ρ is

$$\beta_G = \beta(1 + \rho)$$

and

$$\beta_N = \beta(1 - \rho),$$

with ρ scaled to vary between -1 and 1 (5). The fact that ρ includes not only tonic DA but also phasic DA due to PEs (7) helps to explain why tics can get worse in exciting contexts with positive valence, such as going to Disney World for the first time or during birthdays or holidays, especially when these include presents (148). In this article, however, we focus on the simpler, more routine case in which DA at the time of action selection is determined by tonic DA levels. In that case, it is biologically more realistic to assume that tonic DA affects mostly β_N rather than β_G because of the

Box 1. Continued

high and low affinity of striatal D₂ and D₁ receptors, respectively (see main text). The equation capturing the effect of tonic DA, γ , on action selection therefore can be simplified to

$$P_t(a|s_t) = \frac{e^{\beta G_t(s_t, a) - \beta(1-\gamma)N_t(s_t, a)}}{\sum_{a_i} e^{\beta G_t(s_t, a_i) - \beta(1-\gamma)N_t(s_t, a_i)}}$$

with $\gamma \leq 1$. The basal ganglia have exactly the right anatomy to implement this selection mechanism (Figure 1).

nanomolar DA concentrations, respectively (19–21). Overall, the affinity of striatal D₂ receptors has been estimated to be 67 times higher than that of D₁ receptors (20). Therefore, tonic DA may act predominantly on NoGo (D₂-expressing) MSNs rather than on Go (D₁-expressing) MSNs (22,23). DA reduces the excitability (or gain) of NoGo MSNs (16,24), β_N , thereby suppressing the influence of NoGo values (N) on action selection and thus facilitating action execution (Box 1 and Figure 1). Increased tonic DA therefore invigorates action and amplifies the expression of learned tics (Figure 2A). The evidence for the roles of phasic and tonic DA on action learning and selection, respectively, has been reviewed elsewhere (5–7).

INCREASED PHASIC DA AND ENHANCED REWARD, HABIT, AND TIC LEARNING

Reinforcement Learning

If unmedicated patients with TS have increased phasic DA, they should have enhanced reward learning. Indeed, in an instrumental-learning task with subliminal cues, unmedicated patients with TS learned from gains but not from losses, and antipsychotics reversed those biases, with medicated patients learning from losses but not from gains (25). Patients with Parkinson's disease (PD) exhibited the opposite double dissociation: unmedicated patients learned from losses but not from gains, and patients on levodopa and DA agonists learned from gains but not from losses (25). Better learning from gains and losses in medicated and unmedicated patients with PD, respectively, has also been found with several nonsubliminal tasks (26). The opposite effects in TS and PD provide further evidence for hyperdopaminergia in TS; in addition, they suggest that tics might result from excessive, inappropriate reward learning. Indeed, reward-based motor-skill learning is increased in unmedicated patients with TS and is decreased in patients with TS on antipsychotics (27). Model-based analyses suggested that the increased reward-based learning in unmedicated patients with TS might be due to an increase in the reward representation (r), but the learning rates for the critic and actor were also increased, albeit nonsignificantly (27). Distinguishing between r and learning rates is often difficult (Figure 3). Furthermore, both parameters could capture increased positive-PE signaling, as would be expected with increased phasic DA.

In the aforementioned studies, learning did not appear to be driven by explicit knowledge (25,27). In tasks that rely on explicit knowledge, patients with TS generally do not differ from control subjects. In a deterministic reversal-learning task, patients with TS, with relatively light medication,

performed like healthy control subjects, unless they had comorbid attention-deficit/hyperactivity disorder (ADHD), in which case they performed like patients with ADHD without comorbid TS (28). Nonetheless, patients with TS without comorbid ADHD had an abnormal P2 event-related potential for positive feedback. Patients with TS, mostly unmedicated, also performed like control subjects in the Iowa gambling task (29), in which explicit knowledge also likely plays an important role (30).

Habit Learning

Although the dichotomies between explicit and implicit learning and between goal-directed and habit learning do not entirely overlap, they relate: goal-directed actions are thought to rely on explicit processes, whereas habits are thought to rely on previously acquired, largely implicit stimulus–response associations (31,32). Consistent with the finding that unmedicated patients with TS have increased implicit reward-based learning (25,27), they also have an increased tendency to rely on habit learning relative to goal-directed actions in a reward-based learning context (33). This increased reliance on habits is consistent with the idea that tics are exaggerated motor habits learned by reinforcement learning (RL). Indeed, in unmedicated patients, both tic severity and the tendency to rely on habits correlated with structural connectivity between motor cortices and the posterior putamen (33), which are critical nodes of the motor habit-learning circuit (34–36). An overconnected, overactive habit system therefore might cause increased tendencies for both habits and tics, with tics being a type of motor habit. Indeed, in unmedicated patients, tic severity correlated positively with the tendency to rely on habits (33).

One limitation of this habit-learning study is that patients with TS exhibited a trend for impaired explicit learning of task contingencies (33). The increased reliance on habits therefore could have been caused by difficulties explicitly learning the contingencies to support goal-directed action, as was found in obsessive-compulsive disorder using the same task (37). This concern, however, is partly mitigated by the correlations with connectivity in the habit-learning circuit and by the preserved performance of patients with TS in other tasks involving explicit learning (28,29).

Two studies reported impaired performance in the weather prediction task in patients with TS, which was unlikely to be due to medication or comorbid ADHD (38,39). This task is hypothesized to probe habit learning (40), but it does not include the tests that support confident classification of behaviors as habits (34). These studies also did not disentangle learning from positive versus negative feedback; the impaired

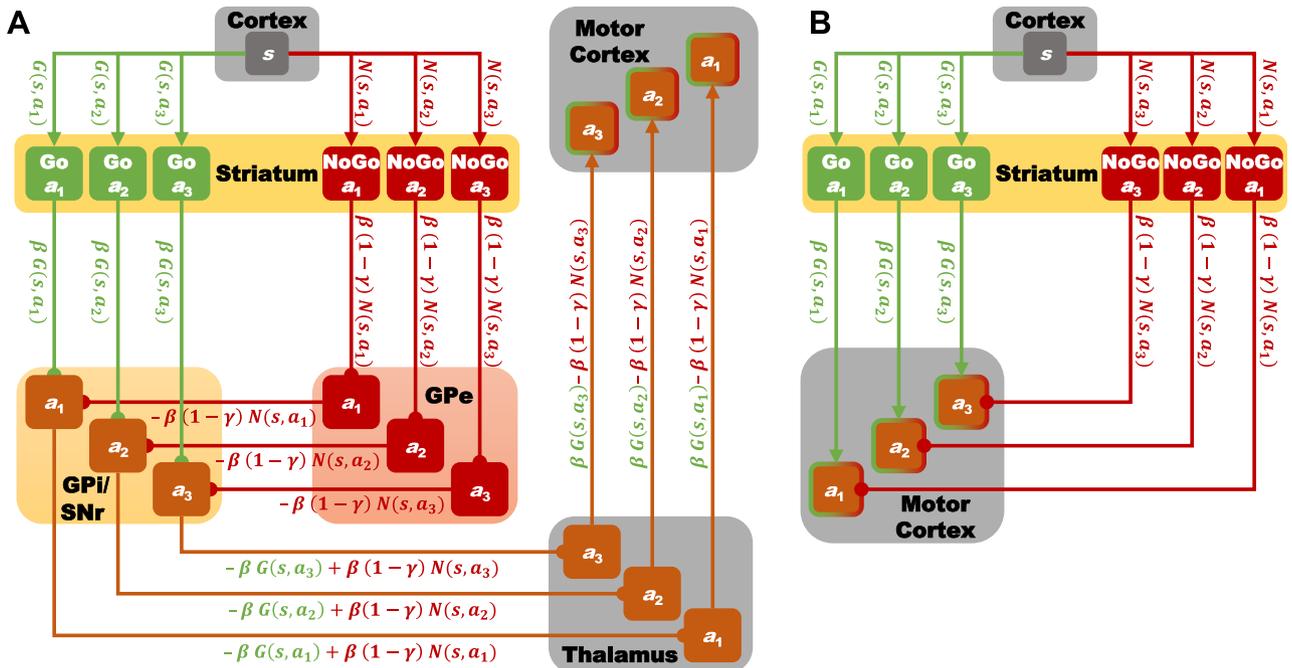


Figure 1. Computational perspectives on action selection by the basal ganglia. **(A)** Reinforcement-learning perspective on the roles of the direct (Go) and indirect (NoGo) basal-ganglia pathways in action selection. The current situation or state, s , is represented in cortex. Three possible actions (a_1 , a_2 , and a_3) are represented; their representation is kept separate in each anatomical region. Striatum contains two populations of neurons: D₁-expressing (Go) and D₂-expressing (NoGo) medium spiny neurons (MSNs), represented in green and red, respectively. Each of these populations contains a representation of the three actions. Go (G) and NoGo (N) values that represent the learned associations between state s and each of the actions are represented in corticostriatal synapses onto Go and NoGo MSNs, respectively. The gain of NoGo MSNs is negatively modulated by tonic dopamine, γ ; the gain of Go MSNs is not modulated by tonic dopamine because of D₁ receptors' lower affinity for DA (see Box 1). The activation of Go and NoGo MSNs therefore is given by $\beta G(s, a_i)$ and $\beta(1-\gamma)N(s, a_i)$, respectively. Mathematically, inhibitory projections (represented by circles) flip the sign of the information (provided that there is intrinsic activity in the target structures, as is the case here). The anatomy of the basal ganglia therefore seems precisely suited to represent the difference between Go and NoGo activations, $\beta G(s, a_i) - \beta(1-\gamma)N(s, a_i)$, in thalamus. Internal variables, structures, and projections related to the direct and indirect pathways are coded in green and red, respectively. Excitatory projections are represented by arrowheads. **(B)** Simplified version of the circuit in **(A)** used in Figure 2. For simplicity and compactness, globus pallidus and thalamus are omitted, so Go and NoGo MSNs are shown directly stimulating and inhibiting motor cortex, respectively. Note from **(A)** that ultimately this is their functional effect. The excitatory and inhibitory connections from Go and NoGo MSNs to motor cortex represent the fact that action values ultimately correspond to a subtraction of the outputs of the Go and NoGo pathways. GPe, globus pallidus external segment; GPI, globus pallidus internal segment; SNr, substantia nigra pars reticulata.

performance in unmedicated patients with TS therefore might have been due to their impairment in learning from negative feedback (25).

Positive Versus Negative Reinforcement

The studies reviewed above suggest that patients with TS exhibit increased learning from rewards (i.e., positive reinforcement). Behavioral models of TS, however, suggest that tics are learned by negative reinforcement, not positive reinforcement, by temporarily decreasing the discomfort associated with premonitory urges (41,42). These ideas can be reconciled because negative reinforcement—reinforcement due to termination or avoidance of aversive stimuli—also relies on positive PEs (6,43,44). Indeed, both termination and avoidance of aversive stimuli elicit phasic DA responses (45–48) that reinforce the action that terminated or avoided the aversive stimulus (47,49). The termination of premonitory urges may similarly elicit phasic DA responses that reinforce tics. If phasic DA, which represents positive PEs, is increased in TS, both positive and negative reinforcement should be increased.

OVERLAPPING NEURAL SUBSTRATES FOR TICS AND HABIT LEARNING AND THEIR RELATION TO HYPERDOPAMINERGIA IN TS

Consistent with the idea that tics are motor habits, the neural substrates of tics prominently include the neural substrates of habit learning. Habit learning relies on the motor cortico–basal ganglia–thalamo–cortical loop, which involves sensorimotor cortices and the putamen (34–36). Much evidence implicates this loop in TS (50–52). Indeed, tic-related activation has been found in all structures of this loop: supplementary motor area (53–56), premotor cortex (53,56,57), sensorimotor cortex (53–57), putamen (53–55,57,58), globus pallidus (54,56), and thalamus (53–56).

Consistent with a hyperdopaminergic account of TS, most findings regarding the motor loop in TS are the opposite of those in PD: 1) several studies (59–63), even if not all (64,65), found increased putamen volumes or gray matter density in TS, whereas putamen volumes are decreased in PD (66); 2) structural connectivity within the motor loop is increased in TS (63) and is decreased in PD (67); 3) resting-state functional connectivity in motor networks is increased in TS (68) and is decreased in PD (69);

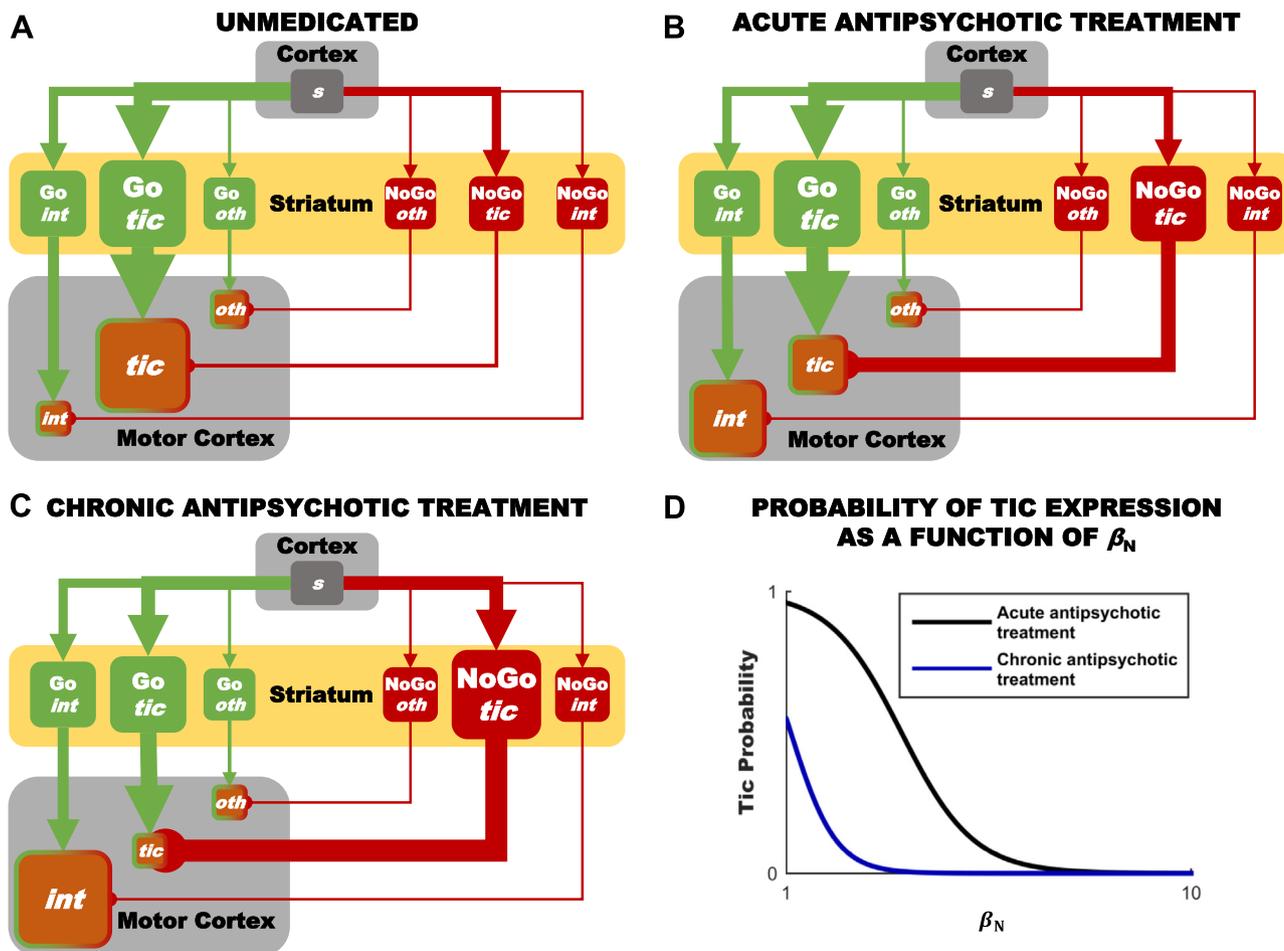


Figure 2. Effects of tonic dopamine (DA) and antipsychotic treatment on Tourette syndrome. Represented in each panel are three actions: an intended action (*int*), a tic (*tic*), and some other weakly supported action (*oth*). Panels (A–C) differ in terms of patient medication status. The figures in each panel follow the representation introduced in Figure 1B. The size of each square and the width of the arrow that departs from it represent the level of neuronal activity. (A) In unmedicated patients with Tourette syndrome, tics may have both 1) strong Go (G) values, learned through aberrant, exaggerated phasic DA responses or through negative reinforcement due to temporary relief of premonitory urges, and 2) strong NoGo (N) values, learned through aversive experiences with tics (e.g., social embarrassment, criticism). However, the high tonic DA suppresses the expression of N values (i.e., it makes β_N very small), so neural activation for the tic in motor cortex is high, making tic expression very likely. (B) Shortly after starting antipsychotic treatment (or reaching a sufficiently high dose), the antipsychotic markedly increases β_N , overweighting N values relative to G values, thereby making the tic less likely to be expressed. (C) Chronic antipsychotic treatment increases N, decreases G, or both; thus, with the same level of D₂ occupancy, and therefore the same increase in β_N , the tic becomes less likely to be expressed than it was shortly after starting antipsychotic treatment [compare with (B)]. For illustrative purposes, the figure shows the case of both increased N values and decreased G values, but the same effect would be obtained with changes in just one or the other. (D) Relation between medication dose and the probability of tic expression with acute treatment (black line) and after chronic treatment has increased N and decreased G (blue line). (Again, the same effect would be obtained by changing only N or G.) Tic suppression is more effective after learning (compare the two lines), which may allow a reduction in dose with no loss of efficacy. If, however, the medication is stopped, tics may return (see the intercept for the blue line): with continued or even reduced medication, β_N amplifies N values relative to G values sufficiently for tics not to be expressed, but when β_N becomes small following antipsychotic withdrawal and the return of high tonic stimulation of postsynaptic D₂ receptors, G values may become preponderant again, causing tics to return. Tic severity, however, may be reduced shortly after the end of treatment relative to before treatment (compare the intercepts for the two lines) because of the partial tic unlearning that occurs during treatment. These curves are intended to capture the phenomena qualitatively, not quantitatively. Details of these simulations are provided in the Supplement. These figures also shed light on why tics often are context dependent (149): tics, like habits, may depend on the situation (s). In fact, the defining characteristic of habits is that they consist of stimulus–response associations (36) (or, more generally, situation–response associations, where a situation can be a complex pattern of external and internal stimuli). Furthermore, the motor loop through the basal ganglia learns precisely those associations (35). Of course, tics may tend to generalize to multiple situations and contexts; (over)generalization, in fact, is a common characteristic of multiple disorders.

4) resting-state spatial covariance analyses showed hyperactivity in premotor and supplementary motor cortices and hypoactivity in thalamus and striatum in TS (70,71), whereas the same analyses showed hypoactivity in premotor and supplementary motor cortices and hyperactivity in thalamus and putamen in PD (72).

MECHANISMS OF ACTION OF ANTIPSYCHOTICS

Fast and Cumulative Effects of Antipsychotics

Antipsychotics have both a fast effect on tic reduction (73,74) and an additional cumulative effect that increases over weeks,

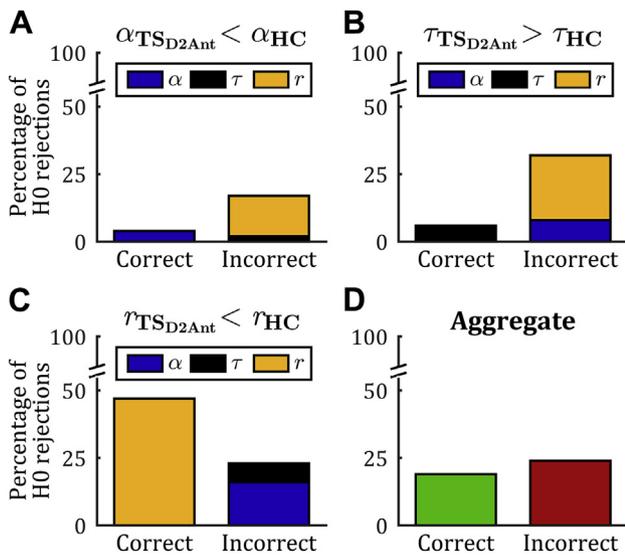


Figure 3. Using models with related parameters can produce misleading results. Many articles use models with parameters that are closely related mathematically—especially some combination of a learning rate, α , a gain, β (or its inverse, a temperature), and a representation of the primary reinforcer, r —without ensuring that the task and fitting procedure used can provide reasonably accurate parameter estimates. The conclusions of those studies can be misleading. To illustrate this point in the context of reinforcement-learning research in Tourette syndrome (TS), we created artificial (simulated) data for a published experiment (100) using, as the generative model, the model that was used in that study to analyze the data. We then applied to our simulated data the same model-fitting procedures and statistical analyses that were applied in the original study to subjects' data. Given that we knew the ground truth by having simulated the data, we could assess the accuracy of the conclusions reached by applying the original model-fitting and statistical procedures. Details about the simulations and analyses are provided in the Supplement. Briefly, the original study used a reinforcement-learning model with three parameters: a learning rate (α), a temperature (τ), and the value of the primary reward (r). [The temperature τ is the inverse of β , that is, $\tau = 1/\beta$. In the original study, the temperature was called β , but here we adopt the more common convention of using τ for the temperature (146).] No model-selection procedures were employed to compare this model with other models, including with reduced models with a subset of these parameters. The main reported result was that patients with TS on antipsychotics (TS_{D2Ant}) had smaller r values than healthy control subjects (HCs) did. In the model, however, α , τ , and r are closely intertwined. To test whether a statistically significant difference in r could be attributed with reasonable confidence to differences in r , rather than in α or τ , we ran three sets of simulations; in each set, the simulated TS_{D2Ant} and HC groups had a very large difference (Cohen's $d = 2.0$) in one of the parameters and did not differ in the other two parameters. The direction of the difference in each of the parameters was such that performance in the TS_{D2Ant} group would be expected to be worse, as was found in the original study. We ran 100 simulations in each set, as if replicating 100 times the original study. In each simulation, we used the model-fitting procedures from the original study and compared the parameters across the two groups using two-tailed unpaired Student's t tests (also as in the original study). We then determined the percentage of simulations in each set in which the null hypothesis (H_0) of no difference between groups for each parameter was rejected: correctly, if the null hypothesis was rejected for the parameter that indeed was different in that set, or incorrectly, if the null hypothesis was rejected for a parameter that did not differ across the groups in that set. (A) Simulations in which the TS_{D2Ant} group had smaller values of α than the HC group, and τ and r did not differ between the groups. Applying the procedures used in the original study resulted in incorrect rejection of the null hypothesis of no differences between r more often than it resulted in correct rejection of the null hypothesis of no differences between α . (B) Simulations in which the TS_{D2Ant}

months, and even years (75–79). The fast effect likely is due to blockade of postsynaptic D_2 receptors, which increases indirect-pathway activity, thereby suppressing tics. Computationally, antipsychotics increase the gain of NoGo MSNs, β_N , thereby amplifying NoGo values ($\beta_N \times N$; Box 1 and Figure 1). Therefore, the learned NoGo values (N) for tics, which result from prior negative experiences with tics—such as embarrassment, criticism, or physical discomfort—get amplified, suppressing tic execution (Figure 2B). The amplified negative (NoGo) values might also include cognitive beliefs (e.g., that tics are bad), which interact with RL processes (80).

The cumulative effects of antipsychotics might be due to gradual biological adaptations. Acute antipsychotic administration increases tonic and phasic DA because of autoreceptor blockade (81–83). Chronic antipsychotic administration, however, reduces tonic and phasic DA (84–86), possibly by inducing depolarization block in DA neurons (86,87). The cumulative effects of antipsychotics therefore might be due to the gradual development of depolarization block. In a hyperdopaminergic animal model, however, acute antipsychotic administration suffices to induce depolarization block (88). Whether that might also occur in TS is unclear; if it does, the gradual improvements with chronic administration might be due to other mechanisms.

An alternative, or additional, explanation for these gradual improvements draws on DA's role in learning. Antipsychotics may cause gradual tic unlearning through mechanisms discussed below that may increase NoGo values (N), decrease Go values (G), or both for tics. Such tic unlearning would explain the tic improvement with continued treatment (Figure 2C), why antipsychotic dose can often be reduced during treatment (75,89) (Figure 2D), and why tics do not always return with full

group had greater values of τ than the HC group, and α and r did not differ between the groups. Applying the procedures used in the original study resulted in incorrect rejection of the null hypothesis of no differences between r more often than it resulted in correct rejection of the null hypothesis of no differences between τ . (C) Simulations in which the TS_{D2Ant} group had smaller values of r than the HC group, and α and τ did not differ between the groups. Applying the procedures used in the original study resulted in correct rejection of the null hypothesis of no differences between r more often than it resulted in incorrect rejections of the null hypotheses for the other parameters. In summary, in all three sets of simulations, the null hypothesis that was rejected most frequently was the one for no differences in r , independent of whether that was the true underlying difference or not. We also calculated the conditional probability of r truly being different given rejection of the null hypothesis for no differences in r ; that probability was .55. Knowing that the original study rejected that null hypothesis therefore provides fairly weak evidence that indeed there was a difference between groups specifically in r because conditional on that rejection, the probability that there was a difference in r (.55) was not much larger than the probability that there was no difference in r (.45). (D) Aggregate percentages of correct and incorrect rejections of the null hypotheses combining the data from the three sets of simulations. The percentage of incorrect rejections was even larger than the percentage of correct rejections. The Supplement contains additional simulations and analyses demonstrating that the problem with the original study (100) was not with the fitting procedure as such but rather with the unsuitability of this task and experimental design to obtain accurate estimates of the highly interrelated parameters in the model. In particular, r strongly correlates negatively with α and positively with τ , which likely explains why differences in those parameters were often captured in r . Other simulations and analyses in the Supplement demonstrate other limitations with the original study and suggest alternative interpretations for its other findings. The Supplement also discusses how these types of problems can be prevented, diagnosed, and overcome.

force—and in some cases may not return at all—following medication withdrawal (74–76) (Figure 2D). Of course, these effects are difficult to disentangle from the natural waxing and waning of tics and from the tic reduction that typically occurs during adolescence and young adulthood (90).

Antipsychotic discontinuation has been associated with a return of tics after a few days (74,89,91), when most of the antipsychotic has left the organism. In two crossover studies, tics increased at the end of the placebo period relative to the end of the preceding treatment period, reaching pretreatment levels in one study (92) but not the other (76). Thus, if tic unlearning occurs during antipsychotic treatment, such unlearning likely is only partial. Computationally, partial tic unlearning is exactly what would be expected: a given increase in N , decrease in G , or both may completely suppress tics during antipsychotic treatment, when β_N is increased, and yet be insufficient to suppress tics after antipsychotic discontinuation, when β_N is reduced because of the high stimulation of postsynaptic D_2 receptors by elevated tonic DA (Figure 2D). In fact, the successful suppression of tics after only partial tic unlearning during treatment, while β_N is elevated, may preclude further tic unlearning because G and N values change only when the action is executed.

Data on illness course after antipsychotic discontinuation are sparse, but tics may tend to gradually worsen (93). Periods of remission of up to 6 months followed by relapse have been reported (75). Again, disentangling these findings from the natural waxing and waning of tics is difficult; nonetheless, gradual worsening of tics following antipsychotic discontinuation could be due to tic learning, which might affect preferentially incompletely unlearned prior tics. Indeed, case reports tend to note the preferential return of prior tics (74,91).

Antipsychotics and Increased NoGo Learning

Long-term depression (LTD) and long-term potentiation (LTP) of corticostriatal synapses onto D_2 MSNs require D_2 -receptor stimulation and lack of stimulation, respectively (14,15). Antipsychotics therefore prevent and facilitate indirect-pathway LTD and LTP, respectively, increasing NoGo learning. Consequently, the NoGo value (N) for tics might increase, making them less likely to occur. Chronic antipsychotic administration induces homeostatic adaptations that counteract the increased NoGo function elicited by D_2 blockade; such chronic administration, however, strengthens a subset of corticostriatal synapses onto D_2 MSNs (94). Which synapses onto D_2 MSNs get strengthened and which get weakened is unclear (94); understanding the biological mechanisms, or possibly the behavioral contingencies, that underlie these differential effects could facilitate better treatments for TS.

Consistent with the idea that antipsychotics may reduce tics in part by increasing NoGo learning, patients with TS on chronic antipsychotic treatment have increased NoGo learning relative to unmedicated patients (25). Increased NoGo learning is also found in unmedicated patients with PD (25,26), healthy subjects under DA depletion (95), and healthy subjects low on striatal tonic DA as assessed indirectly via spontaneous blink rate (96). Low DA therefore also causes increased NoGo learning, likely because of reduced stimulation of postsynaptic D_2 receptors (26).

Antipsychotics and Blunted Go Learning

Antipsychotics impair reward-based learning in animals (97), healthy humans (98,99), and patients with TS (25,100). One study claimed that this impairment is on action selection rather than on learning because 1) the antipsychotic seemed to affect only late, asymptotic behavior, not earlier learning, and 2) in model fits, the affected parameter was the gain for rewards, β_G , not the learning rates for rewards, α_G , or losses, α_N , nor indeed the gain for losses, β_N (99). Such a conclusion, however, seems highly debatable because 1) participants above a median split on serum antipsychotic levels were impaired right from the initial trials, and 2) α and β frequently are not identifiable simultaneously because of their strong mathematical interrelation (Figure 3). Furthermore, this study administered a single high dose of an antipsychotic; an effect on β_G with no effect on β_N seems implausible biologically because a high antipsychotic dose should affect mostly the indirect pathway, not the direct pathway. Finally, studies in animals show that antipsychotics impair reward-based learning over and above possible effects on action selection (97). Consistent with the idea that active-avoidance learning also relies on positive PEs (6,43), antipsychotics also impair active-avoidance learning in healthy humans (98) and animals (101).

Antipsychotic treatment in TS virtually abolishes reward-based learning (25,100) and the effects of rewards on motor-skill learning (27). Notably, antipsychotics do not simply normalize patients' biases: they flip those biases from enhanced reward and blunted punishment learning in unmedicated patients to blunted reward and enhanced punishment learning in medicated patients (25,27,100). This bias reversal may facilitate tic unlearning.

Model-based analyses suggested that antipsychotics have this effect in TS by blunting primary reward, r (not β) (27,100). However, estimating α , β , and r simultaneously in tasks such as the ones in these studies produces unreliable results (Figure 3). Nonetheless, one of these studies showed that antipsychotics blunted neural activation to rewards during learning, with the degree of such blunting explaining the behavioral impairment (100). Overall, these findings suggest that antipsychotics blunt reward-based learning in TS over and above possible effects on action selection; however, they do not license more fine-grained interpretations about the underlying mechanisms (e.g., whether the effect is on primary rewards, PE magnitude, postsynaptic effects of PEs, or another mechanism).

Antipsychotics go beyond blunting Go learning: they cause progressive extinction of previously learned responses (97). Therefore, they should cause gradual tic extinction. Notably, antipsychotics cause response extinction even if the response continues to be reinforced (97). Thus, tics may extinguish even if they continue to alleviate premonitory urges.

The mechanisms by which antipsychotics blunt Go learning are unclear. Chronic antipsychotic administration decreases DA-neuron firing (87,102) and reduces phasic and tonic DA (84–86), which should blunt Go learning and expression. However, acute antipsychotic administration, which increases DA-neuron firing (102) and phasic and tonic DA (81,82), also blunts Go learning (97,99,103). Presynaptic effects therefore are, at best, a partial explanation that applies only to chronic

administration. A postsynaptic explanation, however, seems counterintuitive: antipsychotics block receptors in NoGo MSNs; how can that affect Go learning? We envision three possibilities (that are not mutually exclusive).

First, antipsychotics might not affect the Go pathway: they might just increase NoGo learning and expression, as discussed above. Studies of the effects of antipsychotics on RL typically use probabilistic tasks, which involve Go and NoGo learning for the same stimulus–action pair; increased NoGo learning and expression therefore may counteract Go learning and expression, giving the impression of impaired Go learning. Although this explanation seems likely, it may be insufficient because, in some studies, antipsychotics blunted Go learning without affecting NoGo learning (99,103).

Second, antipsychotics may decrease LTD but not increase LTP in D_2 MSNs. LTD in D_2 MSNs requires both stimulation of D_2 receptors and lack of stimulation of adenosine A_{2A} receptors (14); therefore, antipsychotics are sufficient to block LTD. LTP in D_2 MSNs, however, requires A_{2A} stimulation and lack of D_2 stimulation (14); therefore, antipsychotics alone are insufficient to induce LTP because A_{2A} stimulation is also required. Indeed, antipsychotics abolish LTD but, by themselves, only weakly—if at all—induce LTP in D_2 MSNs (15). In probabilistic tasks, even good actions are occasionally followed by bad outcomes, causing some NoGo learning; impaired LTD in D_2 MSNs would preclude that learning from being “erased” by good outcomes, giving the impression of impaired Go learning. Given that antipsychotics might not increase LTP in D_2 MSNs, they would not improve NoGo learning for bad actions. The behavioral effects therefore would be impaired Go learning but no effect on NoGo learning, as indeed has been found (99,103).

Third, antipsychotics might blunt phasic DA responses via striatal cholinergic tonically active neurons. These neurons pause firing for unpredicted salient positive events, at the same time that DA neurons burst fire (104). The resulting reduction in acetylcholine facilitates DA release during DA-neuron burst firing, when DA neurons are firing at high frequencies, through nicotinic receptors in dopaminergic terminals (104). D_2 blockade in tonically active neurons reduces their firing pauses (105), so it may blunt phasic DA release.

MECHANISMS OF ACTION OF DA AGONISTS

Low doses of some DA agonists, particularly pergolide (106,107), are effective treatments for TS. At low doses, pergolide acts mostly on D_2 autoreceptors, thereby decreasing DA-neuron firing (108,109) and DA levels (110). We are aware of only one RL study with pergolide, in which pergolide dose correlated with reversal errors in patients with PD (111). The doses, however, substantially exceeded those used for TS, so this finding likely is attributable to postsynaptic effects: postsynaptic D_2 agonism should impair learning from negative feedback by inhibiting the indirect pathway. Low doses of pergolide impair other forms of associative learning, probably by blunting phasic DA (112). Furthermore, low doses of cabergoline, another ergot-derived DA agonist with similar binding affinities to

pergolide (113), impair reward-based learning (114). A limitation of these preclinical studies is that they used acute (108–110,114), or at best short-term (112), pergolide administration. Chronic low doses of pergolide may aggravate parkinsonism (115), however, so such chronic administration likely also inhibits DA neurotransmission.

Acute administration of a low dose of pramipexole, another DA agonist, also inhibits DA-neuron firing (116,117) and impairs reward-based learning (118,119). However, a clinical trial of low doses of pramipexole for TS failed (120). The likely reason for this failure is that whereas, acutely, pramipexole inhibits DA-neuron firing (116,117), with chronic administration, DA-neuron firing normalizes (116), and DA levels (121,122) and postsynaptic D_2 stimulation (123) increase. Indeed, the dose (0.5 mg) that impaired RL in control subjects when given acutely (118,119,124) is used, twice daily, to treat early-stage PD (125). Furthermore, in patients with PD with comorbid impulse control disorders using DA agonists chronically, most of whom were on 0.5 to 1 mg of pramipexole, the DA agonist increased reward-based learning (126). The contrasting effects of chronic low doses of pergolide and pramipexole, illustrated by their respective aggravation (115) and amelioration (125) of PD, may relate to their different affinities for the D_3 receptor (Maia and Conceição, unpublished review, February 2017).

MECHANISMS OF ACTION OF OTHER MEDICATIONS

TS is often treated with α_2 agonists, which may decrease both tonic and phasic DA (Maia and Conceição, unpublished review, February 2017). Tetrabenazine, an inhibitor of the vesicular monoamine transporter 2 used to treat TS (127), also decreases both tonic DA (128,129)—possibly after a short-duration increase (128,130)—and phasic DA (130).

COMORBIDITY BETWEEN TS AND ADHD

More than half of patients with TS have comorbid ADHD (131), typically including hyperactivity–impulsivity (i.e., of the combined or hyperactive–impulsive subtypes) (132,133). Conversely, more than 20% of patients with the hyperactive–impulsive subtype of ADHD have comorbid TS, whereas only about half as many with the inattentive subtype do (134). In the general population, TS-like symptoms are also associated with hyperactivity–impulsivity but not with inattention (135). An association between TS and hyperactivity is predicted by our account because increased tonic DA amplifies both tics and the overall tendency for action.

DEVELOPMENTAL COURSE OF TS

Tics tend to increase until periadolescence (around 10 years of age) and then decrease throughout adolescence (136). In rodents, striatal D_1 and D_2 receptors follow exactly that pattern (137). Notably, only male rodents exhibit the peak around periadolescence (138), which might explain the increased prevalence of TS in male individuals. Data in humans are sparser, but striatal D_1 and D_2 receptors and tyrosine hydroxylase activity seem to decrease throughout adolescence (139–141). The course of TS therefore may parallel the course of functional activation of the DA system.

THE CLASSICAL THEORY OF TONIC-PHASIC DA DYSFUNCTION IN TS

A prominent theory suggests that the primary disturbance in TS is an overactive DA transporter, which reduces DA spillage beyond the synapse, thereby causing reduced tonic DA; reduced tonic DA, in turn, would reduce autoreceptor stimulation, which, together with the increased DA available in pre-synaptic terminals due to increased reuptake, would increase phasic DA (3,142). An obvious problem with that theory is that if the main disturbance in TS were an overactive dopamine transporter, then dopamine transporter inhibition—using, for example, psychostimulants or bupropion—should be an excellent treatment for TS, which is not the case. Multiple other lines of evidence, which we review in a recently submitted article, also make that theory unlikely (Maia and Conceição, unpublished review, February 2017).

CONCLUSIONS

Our suggestion that TS involves increased tonic and phasic DA is grounded in an extensively validated computational understanding of the roles of phasic and tonic DA in action learning and selection, respectively, and it provides a coherent, integrated explanation for a broad range of clinical, experimental, and pharmacological findings in TS. The distinction between the roles of increased phasic and tonic DA on tic learning and expression, respectively, also has practical implications for the rational development of new drugs for TS. For example, it suggests that drugs that reduce phasic DA but not tonic DA might be better at preventing learning of new tics than at suppressing existing tics, whereas drugs that reduce tonic DA but not phasic DA might suppress tic expression but not prevent the learning of new tics (which might remain latent for the duration of treatment because of the drug-induced suppression of tic expression).

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REFERENCES

- Leckman JF, Cohen DJ (1983): Recent advances in Gilles de la Tourette syndrome: Implications for clinical practice and future research. *Psychiatr Dev* 1:301–316.
- Singer HS, Butler IJ, Tune LE, Seifert WE, Coyle JT (1982): Dopaminergic dysfunction in Tourette syndrome. *Ann Neurol* 12:361–366.
- Buse J, Schoenfeld K, Münchau A, Roessner V (2013): Neuro-modulation in Tourette syndrome: Dopamine and beyond. *Neurosci Biobehav Rev* 37:1069–1084.
- Singer HS (2013): The neurochemistry of Tourette syndrome. In: Martino D, Leckman JF, editors. *Tourette Syndrome*. New York: Oxford University Press, 276–300.
- Collins AGE, Frank MJ (2014): Opponent actor learning (OpAL): Modeling interactive effects of striatal dopamine on reinforcement learning and choice incentive. *Psychol Rev* 121:337–366.
- Maia TV, Frank MJ (2011): From reinforcement learning models to psychiatric and neurological disorders. *Nat Neurosci* 14:154–162.
- Maia TV, Frank MJ (2017): An integrative perspective on the role of dopamine in schizophrenia. *Biol Psychiatry* 81:52–66.
- Maia TV (2009): Reinforcement learning, conditioning, and the brain: Successes and challenges. *Cogn Affect Behav Neurosci* 9:343–364.
- Schultz W (1997): A neural substrate of prediction and reward. *Science* 275:1593–1599.
- Huys QJM, Tobler PN, Hasler G, Flagel SB (2014): The role of learning-related dopamine signals in addiction vulnerability. *Prog Brain Res* 211:31–77.
- Dayan P (2012): Instrumental vigour in punishment and reward. *Eur J Neurosci* 35:1152–1168.
- Niv Y, Daw ND, Joel D, Dayan P (2007): Tonic dopamine: Opportunity costs and the control of response vigor. *Psychopharmacology (Berl)* 191:507–520.
- Salamone JD, Yohn SE, López-Cruz L, San Miguel N, Correa M (2016): Activational and effort-related aspects of motivation: Neural mechanisms and implications for psychopathology. *Brain* 139:1325–1347.
- Lerner TN, Kreitzer AC (2011): Neuromodulatory control of striatal plasticity and behavior. *Curr Opin Neurobiol* 21:322–327.
- Shen W, Flajolet M, Greengard P, Surmeier DJ (2008): Dichotomous dopaminergic control of striatal synaptic plasticity. *Science* 321:848–851.
- Shen W, Plokin JL, Zhai S, Surmeier DJ (2017): Dopaminergic modulation of glutamatergic signaling in striatal spiny projection neurons. In: Steiner H, Tseng KY, editors. *Handbook of Basal Ganglia Structure and Function*, 2nd ed. (Handbook of Behavioral Neuroscience, vol. 24). Amsterdam: Elsevier, 179–196.
- Robinson DL, Wightman RM (2007): Rapid dopamine release in freely moving rats. In: Michael AC, Borland LM, editors. *Electrochemical Methods for Neuroscience*. Boca Raton, FL: CRC Press, Taylor & Francis Group, 17–34.
- Justice JB (1993): Quantitative microdialysis of neurotransmitters. *J Neurosci Methods* 48:263–276.
- Richfield EK, Penney JB, Young AB (1989): Anatomical and affinity state comparisons between dopamine D1 and D2 receptors in the rat central nervous system. *Neuroscience* 30:767–777.
- Marcellino D, Kehr J, Agnati LF, Fuxe K (2012): Increased affinity of dopamine for D2-like versus D1-like receptors: Relevance for volume transmission in interpreting PET findings. *Synapse* 66:196–203.
- Neve KA, Neve RL (1997): Molecular biology of dopamine receptors. In: Neve KA, Neve RL, editors. *The Dopamine Receptors*. Totowa, NJ: Humana Press, 27–76.
- Dreyer JK, Herik KF, Berg RW, Hounsgaard JD (2010): Influence of phasic and tonic dopamine release on receptor activation. *J Neurosci* 30:14273–14283.
- Surmeier DJ, Carrillo-Reid L, Bargas J (2011): Dopaminergic modulation of striatal neurons, circuits, and assemblies. *Neuroscience* 198:3–18.
- Tritsch NX, Sabatini BL (2012): Dopaminergic modulation of synaptic transmission in cortex and striatum. *Neuron* 76:33–50.
- Palminteri S, Lebreton M, Worbe Y, Gräbi D, Hartmann A, Pessiglione M (2009): Pharmacological modulation of subliminal learning in Parkinson's and Tourette's syndromes. *Proc Natl Acad Sci U S A* 106:19179–19184.
- Wiecki TV, Frank MJ (2010): Neurocomputational models of motor and cognitive deficits in Parkinson's disease. *Prog Brain Res* 183:275–297.
- Palminteri S, Lebreton M, Worbe Y, Hartmann A, Lehericy S, Vidailhet M, et al. (2011): Dopamine-dependent reinforcement of motor skill learning: Evidence from Gilles de la Tourette syndrome. *Brain* 134:2287–2301.

28. Shephard E, Jackson GM, Groom MJ (2016): Electrophysiological correlates of reinforcement learning in young people with Tourette syndrome with and without co-occurring ADHD symptoms. *Int J Dev Neurosci* 51:17–27.
29. Crawford S, Channon S, Robertson MM (2005): Tourette's syndrome: Performance on tests of behavioural inhibition, working memory and gambling. *J Child Psychol Psychiatry* 46:1327–1336.
30. Maia TV, McClelland JL (2004): A reexamination of the evidence for the somatic marker hypothesis: What participants really know in the Iowa Gambling Task. *Proc Natl Acad Sci U S A* 101:16075–16080.
31. Otto AR, Gershman SJ, Markman AB, Daw ND (2013): The curse of planning: Dissecting multiple reinforcement-learning systems by taxing the central executive. *Psychol Sci* 24:751–761.
32. Wood W, Labrecque JS, Lin P-Y, Runger D (2014): Habits in dual process models. In: Sherman JW, Gawronski B, Trope Y, editors. *Dual-Process Theories of the Social Mind*. New York: Guilford, 371–385.
33. Delorme C, Salvador A, Valabrègue R, Roze E, Palminteri S, Vidailhet M, *et al.* (2016): Enhanced habit formation in Gilles de la Tourette syndrome. *Brain* 139:605–615.
34. Balleine BW, O'Doherty JP (2010): Human and rodent homologies in action control: Corticostriatal determinants of goal-directed and habitual action. *Neuropsychopharmacology* 35:48–69.
35. Horga G, Maia TV, Marsh R, Hao X, Xu D, Duan Y, *et al.* (2015): Changes in corticostriatal connectivity during reinforcement learning in humans. *Hum Brain Mapp* 36:793–803.
36. Yin HH, Knowlton BJ (2006): The role of the basal ganglia in habit formation. *Nat Rev Neurosci* 7:464–476.
37. Gillan CM, Pappmeyer M, Morein-Zamir S, Sahakian BJ, Fineberg NA, Robbins TW, de Wit S (2011): Disruption in the balance between goal-directed behavior and habit learning in obsessive-compulsive disorder. *Am J Psychiatry* 168:718–726.
38. Kéri S, Szlobodnyik C, Benedek G, Janka Z, Gáboros J (2002): Probabilistic classification learning in Tourette syndrome. *Neuropsychologia* 40:1356–1362.
39. Marsh R, Alexander GM, Packard MG, Zhu H, Wingard JC, Quackenbush G, Peterson BS (2004): Habit learning in Tourette syndrome: A translational neuroscience approach to a developmental psychopathology. *Arch Gen Psychiatry* 61:1259–1268.
40. Packard MG, Knowlton BJ (2002): Learning and memory functions of the basal ganglia. *Annu Rev Neurosci* 25:563–593.
41. Brandt VC, Beck C, Sajin V, Baaske MK, Bäumer T, Beste C, *et al.* (2016): Temporal relationship between premonitory urges and tics in Gilles de la Tourette syndrome. *Cortex* 77:24–37.
42. Capriotti MR, Brandt BC, Turkel JE, Lee H-J, Woods DW (2014): Negative reinforcement and premonitory urges in youth with Tourette syndrome: An experimental evaluation. *Behav Modif* 38:276–296.
43. Maia TV (2010): Two-factor theory, the actor-critic model, and conditioned avoidance. *Learn Behav* 38:50–67.
44. Seymour B, O'Doherty JP, Koltzenburg M, Wiech K, Frackowiak R, Friston K, Dolan R (2005): Opponent appetitive-aversive neural processes underlie predictive learning of pain relief. *Nat Neurosci* 8:1234–1240.
45. Brischoux F, Chakraborty S, Brierley DI, Ungless MA (2009): Phasic excitation of dopamine neurons in ventral VTA by noxious stimuli. *Proc Natl Acad Sci U S A* 106:4894–4899.
46. Budygin EA, Park J, Bass CE, Grinevich VP, Bonin KD, Wightman RM (2012): Aversive stimulus differentially triggers subsecond dopamine release in reward regions. *Neuroscience* 201:331–337.
47. Navratilova E, Xie JY, Okun A, Qu C, Eyde N, Ci S, *et al.* (2012): Pain relief produces negative reinforcement through activation of mesolimbic reward-valuation circuitry. *Proc Natl Acad Sci U S A* 109:20709–20713.
48. Oleson EB, Gentry RN, Chioma VC, Cheer JF (2012): Subsecond dopamine release in the nucleus accumbens predicts conditioned punishment and its successful avoidance. *J Neurosci* 32:14804–14808.
49. Shumake J, Ilango A, Scheich H, Wetzel W, Ohi FW (2010): Differential neuromodulation of acquisition and retrieval of avoidance learning by the lateral habenula and ventral tegmental area. *J Neurosci* 30:5876–5883.
50. Felling RJ, Singer HS (2011): Neurobiology of Tourette syndrome: Current status and need for further investigation. *J Neurosci* 31:12387–12395.
51. Neuner I, Schneider F, Shah NJ (2013): Functional neuroanatomy of tics. In: Martino D, Cavanna AE, editors. *Advances in Neurochemistry and Neuropharmacology of Tourette Syndrome (International Review of Neurobiology, vol. 112)*. London: Academic Press, 35–71.
52. Worbe Y, Gerardin E, Hartmann A, Valabregue R, Chapin M, Tremblay L, *et al.* (2010): Distinct structural changes underpin clinical phenotypes in patients with Gilles de la Tourette syndrome. *Brain* 133:3649–3660.
53. Bohlhalter S (2006): Neural correlates of tic generation in Tourette syndrome: An event-related functional MRI study. *Brain* 129:2029–2037.
54. Lerner A, Bagic A, Boudreau EA, Hanakawa T, Pagan F, Mari Z, *et al.* (2007): Neuroimaging of neuronal circuits involved in tic generation in patients with Tourette syndrome. *Neurology* 68:1979–1987.
55. Neuner I, Werner CJ, Arrubla J, Stöcker T, Ehlen C, Wegener HP, *et al.* (2014): Imaging the where and when of tic generation and resting state networks in adult Tourette patients. *Front Hum Neurosci* 8:362.
56. Wang Z, Maia TV, Marsh R, Colibazzi T, Gerber A, Peterson BS (2011): The neural circuits that generate tics in Tourette's syndrome. *Am J Psychiatry* 168:1326–1337.
57. Stern E, Silbersweig DA, Chee K-Y, Holmes A, Robertson MM, Trimble M, *et al.* (2000): A functional neuroanatomy of tics in Tourette syndrome. *Arch Gen Psychiatry* 57:741–748.
58. Daubner SC, Le T, Wang S (2011): Tyrosine hydroxylase and regulation of dopamine synthesis. *Arch Biochem Biophys* 508:1–12.
59. Draganski B, Martino D, Cavanna AE, Hutton C, Orth M, Robertson MM, *et al.* (2010): Multispectral brain morphometry in Tourette syndrome persisting into adulthood. *Brain* 133:3661–3675.
60. Ludolph AG, Juengling FD, Libal G, Ludolph AC, Fegert JM, Kassubek J (2006): Grey-matter abnormalities in boys with Tourette syndrome: Magnetic resonance imaging study using optimised voxel-based morphometry. *Br J Psychiatry* 188:484–485.
61. Roessner V, Overlack S, Schmidt-Samoa C, Baudewig J, Dechent P, Rothenberger A, Helms G (2011): Increased putamen and callosal motor subregion in treatment-naïve boys with Tourette syndrome indicates changes in the bihemispheric motor network. *J Child Psychol Psychiatry* 52:306–314.
62. Wen H, Liu Y, Reik I, Wang S, Chen Z, Zhang J, *et al.* (2017): Multimodal multiple kernel learning for accurate identification of Tourette syndrome children. *Pattern Recognit* 63:601–611.
63. Worbe Y, Marrakchi-Kacem L, Lecomte S, Valabregue R, Poupon F, Guevara P, *et al.* (2015): Altered structural connectivity of corticostriato-pallido-thalamic networks in Gilles de la Tourette syndrome. *Brain* 138:472–482.
64. Forde NJ, Zwiers MP, Naaijen J, Akkermans SEA, Openneer TJC, Visscher F, *et al.* (2017): Basal ganglia structure in Tourette's disorder and/or attention-deficit/hyperactivity disorder. *Mov Disord* 32:601–604.
65. Peterson BS, Thomas P, Kane MJ, Scahill L, Zhang H, Bronen R, *et al.* (2003): Basal ganglia volumes in patients with Gilles de la Tourette syndrome. *Arch Gen Psychiatry* 60:415–424.
66. Sterling NW, Lewis MM, Du G, Huang X (2016): Structural imaging and Parkinson's disease: Moving toward quantitative markers of disease progression. *J Park Dis* 6:557–567.
67. Sharman M, Valabregue R, Perlberg V, Marrakchi-Kacem L, Vidailhet M, Benali H, *et al.* (2013): Parkinson's disease patients show reduced cortical-subcortical sensorimotor connectivity. *Mov Disord* 28:447–454.
68. Worbe Y, Malherbe C, Hartmann A, Pelegri-Issac M, Messe A, Vidailhet M, *et al.* (2012): Functional immaturity of cortico-basal ganglia networks in Gilles de la Tourette syndrome. *Brain* 135:1937–1946.
69. Baggio HC, Segura B, Junque C (2015): Resting-state functional brain networks in Parkinson's disease. *CNS Neurosci Ther* 21:793–801.
70. Eidelberg D, Moeller JR, Antonini A, Kazumata K, Dhawan V, Budman C, Feigin A (1997): The metabolic anatomy of Tourette's syndrome. *Neurology* 48:927–933.

71. Pourfar M, Feigin A, Tang CC, Carbon-Correll M, Bussa M, Budman C, *et al.* (2011): Abnormal metabolic brain networks in Tourette syndrome. *Neurology* 76:944–952.
72. Niethammer M, Feigin A, Eidelberg D (2012): Functional neuroimaging in Parkinson's disease. *Cold Spring Harb Perspect Med* 2:a009274.
73. Seignot JN (1961): Un cas de maladie des tics de Gilles de la Tourette guéri par le R-1625. *Ann Méd-Psychol* 119:578–579.
74. Shapiro AK, Shapiro E (1968): Treatment of Gilles de la Tourette's syndrome with haloperidol. *Br J Psychiatry* 114:345–350.
75. Bruun RD, Shapiro AK, Shapiro E, Sweet R, Wayne H, Solomon GE (1976): A follow-up of 78 patients with Gilles de la Tourette's syndrome. *Am J Psychiatry* 133:944–947.
76. George MS, Trimble MR, Robertson MM (1993): Fluvoxamine and sulpiride in comorbid obsessive-compulsive disorder and Gilles de la Tourette syndrome. *Hum Psychopharmacol Clin Exp* 8:327–334.
77. Ghanizadeh A, Haghghi A (2014): Aripiprazole versus risperidone for treating children and adolescents with tic disorder: A randomized double blind clinical trial. *Child Psychiatry Hum Dev* 45:596–603.
78. Neuner I, Nordt C, Schneider F, Kawohl W (2012): Effectiveness of aripiprazole in the treatment of adult Tourette patients up to 56 months. *Hum Psychopharmacol Clin Exp* 27:364–369.
79. Scahill L, Leckman JF, Schultz RT, Katsovich L, Peterson BS (2003): A placebo-controlled trial of risperidone in Tourette syndrome. *Neurology* 60:1130–1135.
80. Doll BB, Jacobs WJ, Sanfey AG, Frank MJ (2009): Instructional control of reinforcement learning: A behavioral and neurocomputational investigation. *Brain Res* 1299:74–94.
81. Imperato A, Chiara GD (1985): Dopamine release and metabolism in awake rats after systemic neuroleptics as studied by trans-striatal dialysis. *J Neurosci* 5:297–306.
82. May LJ, Wightman RM (1989): Effects of D-2 antagonists on frequency-dependent stimulated dopamine overflow in nucleus accumbens and caudate-putamen. *J Neurochem* 53:898–906.
83. Rice ME, Patel JC, Cragg SJ (2011): Dopamine release in the basal ganglia. *Neuroscience* 198:112–137.
84. Cho S, Duchemin A-M, Neff NH, Hadjiconstantinou M (1999): Tyrosine hydroxylase, aromatic L-amino acid decarboxylase and dopamine metabolism after chronic treatment with dopaminergic drugs. *Brain Res* 830:237–245.
85. Feasey-Truger KJ, Earl CD, Alzheimer C, ten Bruggencate G (1995): Stimulus-evoked dopamine overflow in the rat nucleus accumbens is decreased following chronic haloperidol administration: An *in vivo* voltammetric study. *Neurosci Lett* 183:91–95.
86. Lane RF, Blaha CD (1987): Chronic haloperidol decreases dopamine release in striatum and nucleus accumbens *in vivo*: Depolarization block as a possible mechanism of action. *Brain Res Bull* 18:135–138.
87. Grace AA, Bunney BS, Moore H, Todd CL (1997): Dopamine-cell depolarization block as a model for the therapeutic actions of antipsychotic drugs. *Trends Neurosci* 20:31–37.
88. Valenti O, Cifelli P, Gill KM, Grace AA (2011): Antipsychotic drugs rapidly induce dopamine neuron depolarization block in a developmental rat model of schizophrenia. *J Neurosci* 31:12330–12338.
89. Shapiro AK (1973): Treatment of Tourette's syndrome with haloperidol: Review of 34 cases. *Arch Gen Psychiatry* 28:92–97.
90. Bloch MH, Leckman JF (2009): Clinical course of Tourette syndrome. *J Psychosom Res* 67:497–501.
91. Stevens JR, Blachly PH (1966): Successful treatment of the maladie des tics: Gilles de la Tourette's syndrome. *Am J Dis Child* 112:541–545.
92. Gilbert DL, Batterson JR, Sethuraman G, Sallee FR (2004): Tic reduction with risperidone versus pimozide in a randomized, double-blind, crossover trial. *J Am Acad Child Adolesc Psychiatry* 43:206–214.
93. Tourette Syndrome Study Group (1999): Short-term versus longer term pimozide therapy in Tourette's syndrome: A preliminary study. *Neurology* 52:874–877.
94. Sebel LE, Graves SM, Chan CS, Surmeier DJ (2017): Haloperidol selectively remodels striatal indirect pathway circuits. *Neuropsychopharmacology* 42:963–973.
95. Cox SML, Frank MJ, Larcher K, Fellows LK, Clark CA, Leyton M, Dagher A (2015): Striatal D1 and D2 signaling differentially predict learning from positive and negative outcomes. *NeuroImage* 109:95–101.
96. Slagter HA, Georgopoulou K, Frank MJ (2015): Spontaneous eye blink rate predicts learning from negative, but not positive, outcomes. *Neuropsychologia* 71:126–132.
97. Wise RA (2004): Dopamine, learning and motivation. *Nat Rev Neurosci* 5:483–494.
98. Jocham G, Klein TA, Ullsperger M (2014): Differential modulation of reinforcement learning by D2 dopamine and NMDA glutamate receptor antagonism. *J Neurosci* 34:13151–13162.
99. Eisenegger C, Naef M, Linssen A, Clark L, Gandamaneni PK, Müller U, Robbins TW (2014): Role of dopamine D2 receptors in human reinforcement learning. *Neuropsychopharmacology* 39:2366–2375.
100. Worbe Y, Palminteri S, Hartmann A, Vidailhet M, Lehericy S, Pessiglione M (2011): Reinforcement learning and Gilles de la Tourette syndrome. *Arch Gen Psychiatry* 68:1257–1266.
101. Wadenberg M-LG, Hicks PB (1999): The conditioned avoidance response test re-evaluated: Is it a sensitive test for the detection of potentially atypical antipsychotics? *Neurosci Biobehav Rev* 23:851–862.
102. Bunney BS, Grace AA (1978): Acute and chronic haloperidol treatment: Comparison of effects on nigral dopaminergic cell activity. *Life Sci* 23:1715–1727.
103. Pessiglione M, Seymour B, Flandin G, Dolan RJ, Frith CD (2006): Dopamine-dependent prediction errors underpin reward-seeking behaviour in humans. *Nature* 442:1042–1045.
104. Threlfell S, Cragg SJ (2011): Dopamine signaling in dorsal versus ventral striatum: The dynamic role of cholinergic interneurons. *Front Syst Neurosci* 5:11.
105. Kharkwal G, Brami-Cherrier K, Lizardi-Ortiz JE, Nelson AB, Ramos M, Del Barrio D, *et al.* (2016): Parkinsonism driven by antipsychotics originates from dopaminergic control of striatal cholinergic interneurons. *Neuron* 91:67–78.
106. Gilbert DL, Dure L, Sethuraman G, Raab D, Lane J, Sallee FR (2003): Tic reduction with pergolide in a randomized controlled trial in children. *Neurology* 60:606–611.
107. Gilbert DL, Sethuraman G, Sine L, Peters S, Sallee FR (2000): Tourette's syndrome improvement with pergolide in a randomized, double-blind, crossover trial. *Neurology* 54:1310–1315.
108. Carlson JH, Bergstrom DA, Walters JR (1987): Stimulation of both D1 and D2 dopamine receptors appears necessary for full expression of postsynaptic effects of dopamine agonists: A neurophysiological study. *Brain Res* 400:205–218.
109. White FJ, Wang RY (1984): Pharmacological characterization of dopamine autoreceptors in the rat ventral tegmental area: Microiontophoretic studies. *J Pharmacol Exp Ther* 231:275–280.
110. Dethy S, Laute MA, Luxen A, Hildebrand J, Goldman S (1995): Effect of pergolide on endogenous and exogenous L-DOPA metabolism in the rat striatum: A microdialysis study. *J Neural Transm Gen Sect* 101:1–11.
111. Swainson R, Rogers RD, Sahakian BJ, Summers BA, Polkey CE, Robbins TW (2000): Probabilistic learning and reversal deficits in patients with Parkinson's disease or frontal or temporal lobe lesions: Possible adverse effects of dopaminergic medication. *Neuropsychologia* 38:596–612.
112. Breitenstein C, Korsukewitz C, Flöel A, Kretschmar T, Diederich K, Knecht S (2006): Tonic dopaminergic stimulation impairs associative learning in healthy subjects. *Neuropsychopharmacology* 31:2552–2564.
113. Millan MJ, Maiofiss L, Cussac D, Audinot V, Boutin J-A, Newman-Tancredi A (2002): Differential actions of antiparkinson agents at multiple classes of monoaminergic receptor: I. A multivariate analysis of the binding profiles of 14 drugs at 21 native and cloned human receptor subtypes. *J Pharmacol Exp Ther* 303:791–804.

114. Frank MJ, O'Reilly RC (2006): A mechanistic account of striatal dopamine function in human cognition: Psychopharmacological studies with cabergoline and haloperidol. *Behav Neurosci* 120: 497–517.
115. Kellett MW, Steiger MJ (1999): Deterioration in parkinsonism with low-dose pergolide. *J Neurol* 246:309–311.
116. Chernoloz O, El Mansari M, Blier P (2009): Sustained administration of pramipexole modifies the spontaneous firing of dopamine, norepinephrine, and serotonin neurons in the rat brain. *Neuropsychopharmacology* 34:651–661.
117. Piercey MF, Hoffmann WE, Smith MW, Hyslop DK (1996): Inhibition of dopamine neuron firing by pramipexole, a dopamine D3 receptor-preferring agonist: Comparison to other dopamine receptor agonists. *Eur J Pharmacol* 312:35–44.
118. Pizzagalli DA, Evins AE, Schetter EC, Frank MJ, Pajtas PE, Santesso DL, Culhane M (2008): Single dose of a dopamine agonist impairs reinforcement learning in humans: Behavioral evidence from a laboratory-based measure of reward responsiveness. *Psychopharmacology (Berl)* 196:221–232.
119. Santesso DL, Evins AE, Frank MJ, Schetter EC, Bogdan R, Pizzagalli DA (2009): Single dose of a dopamine agonist impairs reinforcement learning in humans: Evidence from event-related potentials and computational modeling of striatal-cortical function. *Hum Brain Mapp* 30:1963–1976.
120. Kurlan R, Crespi G, Coffey B, Mueller-Vahl K, Koval S, Wunderlich G, Pramipexole for TS Trial Investigators (2012): A multicenter randomized placebo-controlled clinical trial of pramipexole for Tourette's syndrome. *Mov Disord* 27:775–778.
121. Castro-Hernández J, Afonso-Oramas D, Cruz-Muros I, Salas-Hernández J, Barroso-Chinea P, Moratalla R, *et al.* (2015): Prolonged treatment with pramipexole promotes physical interaction of striatal dopamine D3 autoreceptors with dopamine transporters to reduce dopamine uptake. *Neurobiol Dis* 74:325–335.
122. Joyce JN, Woolsey C, Ryou H, Borwege S, Hagner D (2004): Low dose pramipexole is neuroprotective in the MPTP mouse model of Parkinson's disease, and downregulates the dopamine transporter via the D3 receptor. *BMC Biol* 2:22.
123. Chernoloz O, El Mansari M, Blier P (2012): Long-term administration of the dopamine D3/2 receptor agonist pramipexole increases dopamine and serotonin neurotransmission in the male rat forebrain. *J Psychiatry Neurosci* 37:113–121.
124. Gallant H, Vo A, Seergobin KN, MacDonald PA (2016): Pramipexole impairs stimulus–response learning in healthy young adults. *Front Neurosci* 10:374.
125. Kieburtt K, Parkinson Study Group PramiBID Investigators (2011): Twice-daily, low-dose pramipexole in early Parkinson's disease: A randomized, placebo-controlled trial. *Mov Disord* 26:37–44.
126. Voon V, Pessiglione M, Brezing C, Gallea C, Fernandez HH, Dolan RJ, Hallett M (2010): Mechanisms underlying dopamine-mediated reward bias in compulsive behaviors. *Neuron* 65:135–142.
127. Thenganatt MA, Jankovic J (2016): Recent advances in understanding and managing Tourette syndrome. *F1000Res* 5.
128. Andersson DR, Nissbrandt H, Bergquist F (2006): Partial depletion of dopamine in substantia nigra impairs motor performance without altering striatal dopamine neurotransmission. *Eur J Neurosci* 24: 617–624.
129. Nunes EJ, Randall PA, Hart EE, Freeland C, Yohn SE, Baqi Y, *et al.* (2013): Effort-related motivational effects of the VMAT-2 inhibitor tetrabenazine: Implications for animal models of the motivational symptoms of depression. *J Neurosci* 33:19120–19130.
130. Owesson-White CA, Roitman MF, Sombers LA, Belle AM, Keithley RB, Peele JL, *et al.* (2012): Sources contributing to the average extracellular concentration of dopamine in the nucleus accumbens. *J Neurochem* 121:252–262.
131. Hirschtritt ME, Lee PC, Pauls DL, Dion Y, Grados MA, Illmann C, *et al.* (2015): Lifetime prevalence, age of risk, and genetic relationships of comorbid psychiatric disorders in Tourette syndrome. *JAMA Psychiatry* 72:325–333.
132. Grados MA, Mathews CA (2008): Latent class analysis of Gilles de la Tourette syndrome using comorbidities: Clinical and genetic implications. *Biol Psychiatry* 64:219–225.
133. Khalifa N, von Knorring A-L (2006): Psychopathology in a Swedish population of school children with tic disorders. *J Am Acad Child Adolesc Psychiatry* 45:1346–1353.
134. Willcutt EG, Nigg JT, Pennington BF, Solanto MV, Rohde LA, Tannock R, *et al.* (2012): Validity of DSM-IV attention deficit/hyperactivity disorder symptom dimensions and subtypes. *J Abnorm Psychol* 121:991–1010.
135. Heym N, Kantini E, Checkley HLR, Cassaday HJ (2014): Tourette-like behaviors in the normal population are associated with hyperactive/impulsive ADHD-like behaviors but do not relate to deficits in conditioned inhibition or response inhibition. *Front Psychol* 5:946.
136. Leckman JF, Zhang H, Vitale A, Lahnin F, Lynch K, Bondi C, *et al.* (1998): Course of tic severity in Tourette syndrome: The first two decades. *Pediatrics* 102:14–19.
137. Wahlstrom D, White T, Luciana M (2010): Neurobehavioral evidence for changes in dopamine system activity during adolescence. *Neurosci Biobehav Rev* 34:631–648.
138. Andersen SL, Rutstein M, Benzo JM, Hostetter JC, Teicher MH (1997): Sex differences in dopamine receptor overproduction and elimination. *NeuroReport* 8:1495–1498.
139. McGeer PL, McGeer EG (1976): Enzymes associated with the metabolism of catecholamines, acetylcholine and GABA in human controls and patients with Parkinson's disease and Huntington's chorea. *J Neurochem* 26:65–76.
140. Montague DM, Lawler CP, Mailman RB, Gilmore JH (1999): Developmental regulation of the dopamine D1 receptor in human caudate and putamen. *Neuropsychopharmacology* 21:641–649.
141. Seeman P, Bzowej NH, Guan HC, Bergeron C, Becker LE, Reynolds GP, *et al.* (1987): Human brain dopamine receptors in children and aging adults. *Synapse* 1:399–404.
142. Singer HS, Szymanski S, Giuliano J, Yokoi F, Dogan AS, Brasic JR, *et al.* (2002): Elevated intrasynaptic dopamine release in Tourette's syndrome measured by PET. *Am J Psychiatry* 159:1329–1336.
143. Barto AG (1995): Adaptive critics and the basal ganglia. In: Houk JC, Davis JL, Beiser DG, editors. *Models of Information Processing in the Basal Ganglia*. Cambridge, MA: MIT Press, 215–232.
144. O'Doherty J, Dayan P, Schultz J, Deichmann R, Friston K, Dolan RJ (2004): Dissociable role of ventral and dorsal striatum in instrumental conditioning. *Science* 304:452–454.
145. Gerfen CR, Surmeier DJ (2011): Modulation of striatal projection systems by dopamine. *Annu Rev Neurosci* 34:441–466.
146. Sutton RS, Barto AG (1998): *Reinforcement Learning: An Introduction*. Cambridge, MA: MIT Press.
147. Frank MJ (2005): Dynamic dopamine modulation in the basal ganglia: A neurocomputational account of cognitive deficits in medicated and nonmedicated parkinsonism. *J Cogn Neurosci* 17:51–72.
148. Leckman JF, Bloch MH, Sukhodolsky DG, Scahill L, King RA (2013): Phenomenology of tics and sensory urges: The self under siege. In: Martino D, Leckman JF, editors. *Tourette Syndrome*. New York: Oxford University Press, 3–25.
149. Himle MB, Capriotti MR, Hayes LP, Ramanujam K, Scahill L, Sukhodolsky DG, *et al.* (2014): Variables associated with tic exacerbation in children with chronic tic disorders. *Behav Modif* 38:163–183.